Investigational Medicinal Chemistry & Pharmacology

Research Article

Open Access

A moderately active crude methanol extract from the whole plant of *Kalanchoe crenata* (Andrews) Haw. (Crassulaceae) strongly potentiated the effect of antibiotics against multidrug-resistant Gram-negative bacteria

Fabrice W. Fokou^{1,2}, Ramelle Ngakam¹, Valaire Y. Matieta¹, Gaelle Kengne Fonkou¹, Stephanie Mapie Tiwa¹, Junior F. Megaptche¹, Paul Nayim¹, Véronique P. Beng², Armelle T. Mbaveng^{1*}, Victor Kuete^{1**}

Abstract

Background: Resistant bacteria develop a high level of resistance to multiple drugs, limiting treatment options and increasing morbidity and mortality. This work was planned to evaluate the antibacterial potential of the methanol extract from the whole plant of *Kalanchoe crenata* (KCW) against multidrug-resistant (MDR) Gram-negative bacteria.

Methods: The minimal inhibitory concentrations (MIC) and the minimal bactericidal concentrations (MBC) of KCW alone, in the presence of an efflux pump inhibitor (EPI) phenylalanine-arginine β -naphthylamide (PA β N), or in the presence of antibiotics were assessed using the broth microdilution method combined with the rapid para-iodonitrotetrazolium chloride (INT) colorimetric technique.

Results: KCW displayed weak antibacterial activities with MIC values ranging from 128 to 1024 μg/mL against 10 of the 15 tested bacterial strains. Moderate antibacterial activities with MIC values ranging from 128-625 μg/mL were recorded against some strains belonging to *Klebsiella pneumoniae, Escherichia coli*, and *Providencia stuartii*. PAβN does not significantly enhance the activity KCW. At MIC/2, KCW potentiated the activity of doxycycline (DOX), levofloxacin (LEV), imipenem (IMI), ciprofloxacin (CIP), ceftriaxone (CRO), and tetracycline (TET) against at least 80% of the MDR bacterial strains tested.

Conclusion: The present study demonstrated that KCW is a moderately active antibacterial agent, but a good efflux pump inhibitor that could potentiate the activity of antibiotics against MDR bacteria over-expressing active efflux pumps.

Keywords: Antibacterial; antibiotic-potentiation; Crassulaceae; efflux pumps; Kalanchoe crenata; multidrug resistance.

Correspondence: *Tel.: +237 676542386; E-mail: <u>armbatsa@yahoo.fr;</u> ORCID: <u>https://orcid.org/0000-0003-4178-4967</u> (Armelle T. Mbaveng); ** Tel.: +237 677355927; E-mail: <u>kuetevictor@yahoo.fr;</u> ORCID: <u>http://orcid.org/0000-0002-1070-1236</u> (Victor Kuete)

¹Department of Biochemistry, Faculty of Science, University of Dschang, Dschang, Cameroon; ²Department of Biochemistry, Faculty of Science, University of Yaoundé I, Yaoundé I, Yaoundé, Cameroon.

Other authors:

E-mail: <u>fabricefokou@yahoo.fr</u> (Fabrice Fokou); *E-mail:* <u>ramellengakam@gmail.com</u> (Ramelle Ngakam); *E-mail:* <u>yvmatieta@yahoo.com</u> (Valaire Y. Matieta); *E-mail:* <u>kengnefonkou15@gmail.com</u> (Gaelle Kengne Fonkou); *E-mail:* <u>stetmapie@gmail.com</u> (Stephanie Mapie Tiwa); *E-mail:* <u>megapfabrice@gmail.com</u> (Junior F. Megaptche); *E-mail:* <u>nayimpaul@yahoo.fr</u> (Paul Nayim); *Email:* <u>v.penlap@yahoo.fr</u> (Véronique P. Beng).

Citation on this article: Fokou FW, Ngakam R, Matieta VY, Kengne Fonkou G, Mapie Tiwa S, Megaptche JF, Nayim P, Beng VP, Mbaveng AT, Kuete V. A moderately active crude methanol extract from the whole plant of Kalanchoe crenata (Andrews) Haw. (Crassulaceae) strongly potentiated the effect of antibiotics against multidrug-resistant Gram-negative bacteria. Investigational Medicinal Chemistry and Pharmacology (2024) 7(1):90; Doi: https://dx.doi.org/10.31183/imcp.2024.00090

Background

Bacterial resistance to antibiotics remains a serious health concern globally [1]. Resistant bacteria develop a high level of resistance to multiple drugs, limiting treatment options and increasing morbidity and mortality. The WHO report attributes 45% of deaths in Africa to multidrug-resistant (MDR) bacteria [1]. It is estimated that between 2015 and 2050, bacterial infections and antimicrobial resistance will have caused approximately 1.3 million deaths in Europe [1]. More than 2.8 million people in the United States die from antibiotic-resistant infections each year [1]. Despite the great diversity of antibiotics used clinically, resistance of bacteria to all classes has been observed to date. This resistance causes a huge financial burden in all countries. For example, the Center for Diseases Control and Prevention (CDC) determined that treating six alarming antibiotic-resistance threats accounts for more than \$4.6 billion USD in healthcare costs annually. These bacteria include vancomycin-resistant Enterococcus, carbapenem-resistant Acinetobacter, methicillin-resistant Staphylococcus aureus (MRSA), carbapenem- and extended-spectrum cephalosporinresistant Enterobacteria, and MDR Pseudomonas aeruginosa [1, 2]. The continuous search for new drugs capable of counteracting various forms of resistance remains therefore a priority for researchers around the world. These new antimicrobials should also provide an advantage related to their lower toxicity compared to existing ones. Medicinal plants are an undeniable source of lowtoxicity drugs that can help in the fight against human diseases [3-8]. Their pharmacological activities against resistant phenotypes of bacteria, parasites, or even cancer cells have now been demonstrated [9-25]. Numerous scientific publications have demonstrated over the past two decades the strong capacity of African medicinal plants and their constituents to hinder the growth of these cell forms that are harmful to animals and humans [26-29]. The intensification of this research will make it possible to afford a good number of bioactive substances that can satisfactorily pass the various clinical phases to have new drugs that are more effective and less harmful. Thus, we were interested in this work, to determine the antibacterial potential of Kalanchoe crenata (Andrews) Haw. (Crassulaceae) on MDR phenotypes. It is a succulent flowering plant native to Madagascar and is commonly known as Kalanchoe, Mother of millions, Never die, Dog's liver, Orange forest kalanchoe. Kalanchoe crenata is present in tropical Africa, from Kenya to Tanzania, Uganda, Burundi, Central African Republic, Rwanda, Zaire, Guinea, Sierra Leone, Angola, but also in South Africa [30]. The plant is traditionally used for the treatment of headache, general debility, dysentery, smallpox and convulsion, earache, wounds and sores, but also as a sedative and a remedy for chronic cough [31]. The plant previously displayed cytotoxic activity towards PC212 human mesothelioma cells, A549 human non-small cell lung cancer (NSCLC) cells, HepG2 hepatocarcinoma cells, MCF-7 breast adenocarcinoma cells, and DLD-1 colorectal adenocarcinoma cell lines [32], analgesic and anticonvulsant effects [33], antihyperglycemic activity [34], and nephroprotective effects [35]. The antibacterial potential of this plant has previously been documented on Pseudomonas aeruginosa, Klebsiella pneumoniae, Bacillus subtilis, Shigella flexneri, Escherichia coli, and Staphylococcus aureus [36].

Methods

Plant mateial and extraction

The whole plant of *Kalanchoe crenata* was collected in Dschang, West Region of Cameroon, in December 2022. The plant was identified at the National Herbarium of Cameroon (HNC) under the identification number 35196/HNC. The whole plant was air-dried and then ground. The powder obtained was macerated in 95% methanol for 48 hours, then the mixture was filtered using Whatman N°1. The resulting filtrate was concentrated under a vacuum (BÜCHI R-200) at 65°C at reduced pressure. The crude extract was completely dried in an oven at 40°C. The obtained crude extract (botanical, KCW) was kept at 4°C till further use.

Chemicals and culture media

The efflux pump inhibitor (EPI), the antibiotics, and bacterial growth revelator among others were purchased from Sigma-Aldrich (St. Quentin Fallavier, France). The EPI used was phenylalanine-arginine β -naphthylamide (PA β N). para-iodonitrotetrazolium chloride \geq 97% (INT) was used as the bacterial growth indicator. Dimethyl sulfoxide (DMSO) served to solubilize the botanical. The antibiotics used include doxycycline (DOX), levofloxacin (LEV), imipenem (IMI), ciprofloxacin (CIP), ampicillin (AMP), ceftriaxone (CRO), tetracycline (TET), and vancomycin (VAN). The culture media used were Mueller Hinton Agar (MHA), for the activation of bacterial strains and isolates, Mueller Hinton Broth (MHB), for antibacterial testing, and Eosin methylene blue (EMB), used as a specific and differential culture medium to confirm the purity of bacterial strains [37-39].

Bacterial species

The Gram-negative bacteria tested included both reference strains and clinical isolates of *Escherichia coli* (ATCC10536, AG102, and AG100), *Klebsiella pneumoniae* (ATCC11296, KP55, and K2), *Pseudomonas aeruginosa* (PA01 and PA124), *Enterobacter aerogenes* (EA3, EA298, and EA27), and *Providencia stuartii* (ATCC29916, PS2636, and NEA16). Their bacterial features were previously reported [37, 39-51]. *Escherichia coli* (AG102, and AG100), *Klebsiella pneumoniae* (KP55, and K2), *Enterobacter aerogenes* (EA3, EA298, and EA27), and *Providencia stuartii* (PS2636, and NEA16) are clinical bacterial strains over-expressing AcrAB-TolC efflux pumps while *Pseudomonas aeruginosa* PA124 over-expressed MexAB-OprM pumps [51-54].

Determination of minimal inhibitory and bactericidal concentrations

The minimal inhibitory concentrations (MIC) and the minimal bactericidal concentrations (MBC) of KCW and antibiotics alone, in the presence of PA β N (EPI) were determined using the combined microbroth dilution and rapid INT colorimetric method as previously described in similar experimental conditions [37, 50, 52, 54-60]. Each experiment was repeated three times in triplicate.

Determination of the antibiotic-potentiating effects of KCW

The effects of the association of KCW at the sub-inhibitory concentrations of MIC/2 and MIC/4 with antibiotics (DOX, LEV, IMI, CIP, AMP, CRO, TET, and VAN) were determined against the MDR bacteria using the rapid INT colorimetric method as previously described in the similar experimental conditions [37-39,

43]. Antibiotic-resistance modulating factor (AMF) was calculated as the ratio of the MIC of the antibiotic alone versus MIC in combination with the plant extract. The potentiation effect was considered for AMF ≥ 2 [61].

Interpretation of antibacterial data

The General interpretation of the antibacterial activity of botanicals was performed according to the cutoff values established by Kuete [62] as follows: significant activity (MIC <100 µg/mL), moderate (100 <MIC \leq 625 µg/mL), and weak (MIC> 625 µg/mL). For Enterobacteria, the following specific cutoff points were used: outstanding activity (MIC ≤8 µg/mL), excellent activity (8 < MIC ≤64 µg/mL), very good activity (64 < MIC ≤128 µg/mL), good activity (128 < MIC \leq 256 µg/mL), average activity (256 < MIC \leq 512 μ g/mL), weak activity (512 < MIC ≤1024 μ g/mL), and not active (MIC values >1024 µg/mL) [63]. For Pseudomonas aeruginosa, the following scales were applied: outstanding activity when MIC \leq 32 μ g/mL; excellent activity when 32 < MIC ≤ 128 μ g/mL; very good activity when 128 < MIC \leq 256 µg/mL; good activity when 256 < MIC \leq 512 µg/mL, average activity when 512 < MIC \leq 1024 µg/mL, weak activity or not active when MIC values >1024 µg/mL [64]. Bactericidal activities are considered when the ratios MBC/MIC are below or equal to 2; MBC/MIC ratios above 2 define the bacteriostatic activities [65-68].

Results

The crude extract from the whole plant of Kalanchoe crenata displays moderate to weak antibacterial effects

The MIC and MBC were determined on a panel of 15 bacterial strains and the results are shown in Table 1. It appears that KCW displayed moderate to weak antibacterial activities with MIC values ranging from 128 to 1024 μ g/mL against 10 of the 15 tested bacterial strains. Moderate antibacterial activities of KCW with MIC values ranging from 128-625 μ g/mL were recorded against *Klebsiella pneumoniae* K2 and ATCC 11296, *Escherichia coli* ATCC10536, and *Providencia stuartii* PS2636 and ATCC 29916. The MBC values were generally above 1024 μ g/mL. The MIC values of the reference antibiotic, IMI, varied from <4 to 128 μ g/mL. This drug generally displayed bacteriostatic effects against the tested bacteria, with most of the MBC/MIC ratios above 2.

 $PA\beta N$ did not enhance the activity of the crude extract from the whole plant of Kalanchoe crenata

To check whether the bacterial efflux pumps are involved in the resistance of the tested bacteria to KCW, the MIC values of the botanical were determined in the presence of the EPI (PA β N). The results are shown in Table 2. In the presence of PA β N, the activity of KCW generally did not change, and only a 2-fold increase was observed on 18% (2/11) of the tested bacteria. This is a clear indication that KCW and its constituents are not the substrates of the bacterial efflux pumps.

The crude extract from the whole plant of Kalanchoe crenata significantly potentiated the activity of antibiotics

KCW at MIC/2 and MIC/4, was combined with antibiotics, and tested against ten bacterial species, including *E. aerogenes* EA3 and EA27, *P. stuartii* PS2636 and NAE16, *E. coli* ATCC10536 and AG100, *P. aeruginosa* PA01 and PA121, and *K. pneumoniae* KP2 and KP55. The results are shown in Table 3. It appears that the antibiacterial activities of the antibiotics increased in the presence of the extracts of KCW at MIC/2 and MIC/4, with the AMF ranging from 2- to 128-fold in most of the cases. At MIC/2, KCW potentiated the activity of LEV, IMI, DOX, CIP, VAN, CRO, and TET against at least 80% of the MDR bacterial strains tested. At MIC/4, KCW also potentiated the activity of DOX, CIP, VAN, and CRO against at least 70% of the MDR bacterial strains tested.

Discussion

African medicinal plants have been shown to be efficient sources of antibacterial agents against both drug sensitive and MDR phenotypes [50, 56, 69-84]. The search for novel antibacterial agents should involve the use of MDR strains. In the present study, several MDR bacteria have been used. This could be evidenced by the increase in the antibacterial activity of the reference drug, IMI in the presence of PABN (Table 2). In effect, PABN has been identified as the efflux pumps inhibitor of AcrAB-TolC in Enterobacteria and MexAB-OprM in Pseudomonas aeruginosa [40]. The botanical from Kalanchoe crenata (KCW) displayed moderate to weak antibacterial activity according to Kuete [62]. However, in Enterobacteria very good activity (64 < MIC ≤128 µg/mL) [63] of KCW was obtained against K. pneumoniae ATCC11296 while average activity (256 < MIC ≤512 µg/mL) [63] was recorded against K. pneumoniae K2, Escherichia coli ATCC10536, and Providencia stuartii PS2636 and ATCC29916 (Table 1). Average antibacterial activity (512 < MIC \leq 1024 µg/mL) [64] was obtained against Pseudomanas aeruginosa PA124. The inhibitory activities of KCW against the other bacterial strains were rather weak or considered not active. The poor antimicrobial activity of this plant has previously been reported against Pseudomonas aeruginosa, Klebsiella pneumoniae, Bacillus subtilis (MIC of 8 mg/mL), Shigella flexneri (MIC of 32 mg/mL), Escherichia coli (MIC of 64 mg/mL), and Staphylococcus aureus (MIC of 128 mg/mL) [36]. These data corroborate the results obtained in the present study, confirming that this plant is a moderately active antibacterial agent, as the overall activity was rather moderate.

It has been suggested that if an antibacterial agent potentiates the activity of at least 70% of antibiotics against at least 70% of the tested bacteria over-expressing active efflux pumps, it should be considered a potential EPI [85, 86]. In the present study, KCW at MIC/2, potentiated the activity of 7/8 (88%) antibiotics (LEV, IMI, DOX, CIP, VAN, CRO, and TET) against at least 80% of the MDR bacterial strains tested (Table 2). Therefore, this botanical is an an EPI.

Table 1. Antibacterial activities of crude methanol extract from the whole plant of Kalanchoe crenata.

| Bacterial strains | Tested samples, MIC and MBC (MIC and MBC in μg/mL), and their ratio | | | | | | | | |
|------------------------|---|------|-------------|-----|-----|----|--|--|--|
| | Botanical | | · · · · · · | ATB | | | | | |
| | KCW | | | IMI | | | | | |
| | MIC | MBC | R | MIC | MBC | R | | | |
| Pseudomonas aeruginosa | | | | | | | | | |
| PA01 | >2048 | - | nd | 16 | 64 | 4 | | | |
| PA121 | >2048 | - | nd | 16 | 128 | 8 | | | |
| PA124 | 1024 | - | nd | 4 | 32 | 8 | | | |
| Klebsiella pneumoniae | | | | | | | | | |
| K2 | 512 | - | nd | <4 | 64 | 16 | | | |
| KP55 | >2048 | - | nd | <4 | 64 | 16 | | | |
| ATCC 11296 | 128 | - | nd | <4 | <4 | 1 | | | |
| Escherichia coli | | | | | | | | | |
| AG100 | 1024 | - | nd | 8 | 64 | 8 | | | |
| AG102 | >2048 | - | nd | 16 | 64 | 4 | | | |
| ATCC10536 | 512 | - | nd | 8 | 64 | 8 | | | |
| Providencia stuartii | | | | | | | | | |
| PS2636 | 512 | 1024 | 2 | 8 | 64 | 8 | | | |
| NEA16 | 2048 | - | nd | 8 | 64 | 8 | | | |
| ATCC 29916 | 512 | - | nd | <4 | 16 | 4 | | | |
| Enterobacter aerogenes | | | | | | | | | |
| EA3 | 2048 | - | nd | 8 | 64 | 8 | | | |
| EA27 | >2048 | - | nd | 8 | 16 | 2 | | | |
| EA298 | 2048 | - | nd | 8 | 8 | 1 | | | |

KCW: crude methanol extract from the whole plant of Kalanchoe crenata; R: MBC/MIC ratio; (-): > 2048 or inactive; (nd): not determined; MIC: minimal inhibitory concentration; MBC: minimal bactericidal concentration; IMI: imipenem; ATB: Antibiotic.

Table 2. Minimal inhibitory concentrations of the crude methanol extract from the whole plant of Kalanchoe crenata in the presence of PAβN.

| Bacterial strains | Tested samples, MIC in the absence or presence of EPI (in μg/mL), and their ratio | | | | | | | | |
|------------------------|---|----------|---|-----------|----------|----|--|--|--|
| | Botanical | | | ATB | | | | | |
| | KCW | | | IMI | | | | | |
| | MIC alone | ΜΙC+ΡΑβΝ | R | MIC alone | ΜΙC+ΡΑβΝ | R | | | |
| Pseudomonas aeruginosa | | | | | | | | | |
| PA01 | 2048 | 2048 | 1 | 16 | 16 | 1 | | | |
| PA121 | 2048 | 2048 | 1 | 16 | 2 | 8 | | | |
| Klebsiella pneumoniae | | | | | | | | | |
| K2 | 512 | 256 | 2 | <4 | 1 | <4 | | | |
| KP55 | 2048 | 2048 | 1 | <4 | 4 | <4 | | | |
| Escherichia coli | | | | | | | | | |
| AG100 | 2048 | 2048 | 1 | 8 | 2 | 4 | | | |
| AG102 | 2048 | 2048 | 1 | 16 | 2 | 8 | | | |
| ATCC10536 | 2048 | 2048 | 1 | 8 | 1 | 8 | | | |
| Providencia stuartii | | | | | | | | | |
| PS2636 | 2048 | 2048 | 1 | 8 | 4 | 2 | | | |
| NEA16 | 2048 | 1024 | 2 | 8 | 8 | 1 | | | |
| Enterobacter aerogenes | | | | | | | | | |
| EA3 | 2048 | 2048 | 1 | 8 | 4 | 2 | | | |
| EA27 | 2048 | 2048 | 1 | 8 | <1 | >8 | | | |

KCW: crude methanol extract from the whole plant of Kalanchoe crenata; IMI: imipenem; R: MIC alone vs MIC with PAβN ratio; MIC alone: Minimal Inhibitory Concentration of the sample alone; MIC+PAβN: Minimal Inhibitory Concentration of the sample in the presence of PAβN; ATB: Antibiotic

Table 3. MICs (µg/mL) of antibiotics of the crude methanol extract from the whole plant of Kalanchoe crenata.

| ATB | Extract | Bacterial strains, MIC of antibiotics in the presence of extract and Antibiotic-resistance modulating factor (AMF) | | | | | | | | | | PSP (%) |
|-----|-------------------|--|----------------|---------------|-------------|--------------|---------------|----------|---------------|----------|-------------|---------|
| | concent ration | E. aerogenes P. stuartii | | E. coli | | | P. aeruginosa | | K. pneumoniae | | - `` | |
| | | EA3 | EA27 | PS2636 | NEA16 | ATCC10536 | AG100 | PA01 | PA121 | KP2 | KP55 | _ |
| LEV | 0 | 1 | 0.5 | 0.25 | 0.5 | 0.25 | 0.25 | 0.5 | 0.5 | 0.5 | 0.5 | |
| | MIC/2 | <0.0625 (16) | <0.0625 (8) | <0.0625 (4) | <0.0625 (8) | < 0.0625(4) | 0.25(1) | 0.5(1) | 0.25(2) | 0.25 (2) | 0.125(4) | 80 |
| | MIC/4 | 0.5(2) | <0.0625 (8) | <0.0625 (4) | <0.0625 (8) | 0.125(2) | 0.25(1) | 1(1) | 1(1) | 0.5(1) | 0.5(1) | 50 |
| IMI | 0 | 8 | 8 | 8 | 8 | 8 | 8 | 16 | 16 | <4 | 4 | |
| | MIC/2 | 1/2 (16) | 1 (8) | 4 (2) | 8(1) | <1/2 (16) | 2 (4) | 4 (4) | 4 (4) | 2 (2) | 1/16 (64) | 90 |
| | MIC/4 | 8 (1) | 2 (4) | 8 (1) | 8(1) | <1/2 (16) | 2 (4) | 16 (1) | 8 (2) | 8 (0.5) | 1/16 (64) | 50 |
| DOX | 0 | 1 | 0.5 | 8 | 8 | 1 `´ | 0.5 | 0.5 | 1`´ | 2`´ | 8 `́ | |
| | MIC/2 | 0.125(8) | 0.0625 (8) | <0.0625 (128) | 0.5(16) | < 0.0625(16) | 0.25(2) | 0.25(2) | 1(1) | 4(0.5) | 1(8) | 80 |
| | MIC/4 | 0.125(8) | 0.25(2) | <0.0625 (128) | 1(8) | < 0.0625(16) | 0.25(2) | 0.25(2) | 1(1) | 4(0.5) | 2(4) | 80 |
| IP | 0 | 1 | 0.25 | 0.25 | 8 | 0.25 | 0.5 | 1 | 0.5 | 0.5 | 0.5 | |
| | MIC/2 | 1(1) | 0.5(0.5) | <0.0625 (4) | 1(8) | < 0.0625(4) | 0.25(2) | 0.25(4) | 0.25(2) | 0.5(1) | <0.0625 (8) | 70 |
| | MIC/4 | 1(1) | 1(0.25) | 0.125(2) | 1(8) | < 0.0625(4) | 0.25(2) | 0.5(2) | 0.5(1) | 0.5(1) | 0.25(2) | 60 |
| VAN | 0 | 64 | 2`́ | 2 | 8 | 2 | 256 | 2 | 2 | >64 | >64 | |
| | MIC/2 | 2 (32) | 1(2) | 1(2) | 8(1) | <0.5(4) | 32(8) | 1(2) | 1 (2) | 1(64) | <0.5 (128) | 90 |
| | MIC/4 | 4(16) | 1(2) | 1(2) | 8(1) | 1(2) | 32(8) | 2(1) | 1 (2) | 8(8) | <0.5 (128) | 80 |
| MP | 0 | >256 | >256 | >256 | >256 | >256 | >256 | >256 | >256 | >256 | >256 | |
| | MIC/2 | >256(1) | <2(128) | >256(1) | 256(1) | >256(1) | >256(1) | >256(1) | 32(8) | >256 (1) | 256(1) | 20 |
| | MIC/4 | >256(1) | 8(32) | >256(1) | >256(1) | >256(1) | >256(1) | >256(1) | >256(1) | >256 (1) | >256(1) | 10 |
| ET | 0 | 8 | >8 | 0.25 | >8 | 4 | 2 | 0.25 | 1 | >8 | >8 | |
| | MIC/2 | <0.0625 (128) | < 0.0625 (128) | <0.0625 (4) | 1(8) | 0.5(8) | 2(1) | 0.125(2) | <0.0625 (16) | >8(1) | 1(8) | 80 |
| | MIC/4 | 0.25(32) | <0.0625 (128) | <0.0625 (4) | 8(1) | 4(1) | 2(1) | 0.25(1) | <0.0625 (16) | >8(1) | 8(1) | 40 |
| RO | 0 | 32 | 32 | 64 | 256 | 8 | 32 | 8 | 16 | 8 | 64 | |
| | MIC/2 | 16(2) | <2(16) | 64(1) | 32(8) | <2 (4) | 8(4) | 4(2) | <2(8) | 1(8) | 16(4) | 80 |
| | MIC/4 | 32(1) | 16(2) | 64(1) | 128(2) | <2 (4) | 8(4) | 8(1) | 8(2) | 2(4) | 32(2) | 70 |

MIC: Minimal Inhibitory Concentration; (): AMF: Antibiotic-resistance modulating Factor; PSP (%): percentage of strain where potentiation effect was observed; ATB: Antibiotic; LEV: Levofloxacin; VAN: Vancomycin; CIP: Ciprofloxacin; TET: Tetracycline; DOX: Doxycycline; IMI: Imipenem; CRO: Ceftriaxone; AMP: Ampicillin.

Conclusion

In the present study, the moderate antibacterial potential of the methanol extract from the whole plant of *Kalanchoe crenata* was demonstrated. It was also demonstrated that the botanical from this plant is an efflux pump inhibitor and therefore could potentiate the activity of antibiotics against MDR bacteria over-expressing active efflux pumps. Further study will be performed to identify phytochemicals responsible for the inhibition of the bacterial efflux pumps. Also, toxicological assessments of the botanical will be performed to evaluate its safety.

Abbreviations

AMP: ampicillin ATCC: American-Type Culture Collection CDC: Center for Diseases Control and Prevention CIP: ciprofloxacin CRO: ceftriaxone DMSO: Dimethyl sulfoxide DOX: doxycycline EMB: Eosin methylene blue EPI: efflux pump inhibitor HNC: National Herbarium of Cameroon IMI: imipenem INT: para-lodonitrotetrazolium chloride LEV: levofloxacin MBC: minimal bactericidal concentration MDR: multidrug-resistant MHA: Mueller Hinton Agar MHB: Mueller Hinton MIC: minimal inhibitory concentrations PA β N: phenylalanine-arginine β -naphthylamide TET: tetracycline VAN: vancomycin ZJB: methanol extract from the whole plant of Kalanchoe crenata

Authors' Contribution

FF, RN, VYM, GKF, SMT, JFM, and PN carried out the study; ATM and VK supervised the study; All authors read and approved the final version of the manuscript.

Acknowledgments

The authors are grateful to the Cameroon National Herbarium for identifying the plant.

Conflict of interest

The authors declare no conflict of interest.

Article history:

Received: 17 June 2023 Received in revised form: 10 August 2023 Accepted: 22 August 2023 Available online: 22 August 2023

References

- Fongang H, Mbaveng AT, Kuete V. 2023. Chapter One Global burden of bacterial infections and drug resistance. Advances in Botanical Research, 106:1-20. https://doi.org/10.1016/bs.abr.2022.08.001.
- CDC. 2021. Partners Estimate Healthcare Cost of Antibiotic-resistant Infections. https://www.cdc.gov/drugresistance/solutions-initiative/stories/partnership-estimateshealthcare-cost.html.
- Poumale HMP, Hamm R, Zang Y, Shiono Y, Kuete V. 2013. 8 Coumarins and Related Compounds from the Medicinal Plants of Africa. In: *Medicinal Plant Research in Africa.* edn. Edited by Kuete V. Oxford: Elsevier, pp. 261-300.
- Omosa LK, Midiwo JO, Kuete V, 2017. Chapter 19 Curcuma longa. In: Medicinal Spices and Vegetables from Africa. edn. Edited by Kuete V: Academic Press. pp. 425-435.
- Mbaveng AT, Hamm R, Kuete V. 2014. 19 Harmful and protective effects of terpenoids from african medicinal plants. In: *Toxicological Survey of African Medicinal Plants*. edn. Edited by Kuete V: Elsevier. pp. 557-576.
- Mbaveng AT, Zhao Q, Kuete V. 2014. 20 Harmful and protective effects of phenolic compounds from african medicinal plants. In: *Toxicological Survey of African Medicinal Plants*. edn. Edited by Kuete V: Elsevier. pp. 577-609.
- Kuete V. 2014. 21 Health Effects of Alkaloids from African Medicinal Plants. In: Toxicological Survey of African Medicinal Plants. edn. Edited by Kuete V: Elsevier. pp. 611-633.
- 8. Kuete V. 2014. Toxicological survey of African medicinal plants. Oxford: Elsevier.
- Boyom FF, Ngouana V, Zollo PH, Menut C, Bessiere JM, Gut J, Rosenthal PJ. 2003. Composition and anti-plasmodial activities of essential oils from some Cameroonian medicinal plants. *Phytochemistry*. 64(7):1269-1275.
- Kuete V, Wiench B, Alsaid MS, Alyahya MA, Fankam AG, Shahat AA, Efferth T. 2013. Cytotoxicity, mode of action and antibacterial activities of selected Saudi Arabian medicinal plants. *BMC Complement Altern Med.* 13:354.
- Zofou D, Tene M, Ngemenya MN, Tane P, Titanji VP. 2011. *In vitro* antiplasmodial activity and cytotoxicity of extracts of selected medicinal plants used by traditional healers of Western cameroon. *Malar Res Treat*. 2011:561342.
- Kuete V, Wiench B, Hegazy ME, Mohamed TA, Fankam AG, Shahat AA, Efferth T. 2012. Antibacterial activity and cytotoxicity of selected Egyptian medicinal plants. *Planta Med.* 78(2):193-199.
- Voukeng IK, Beng VP, Kuete V. 2016. Antibacterial activity of six medicinal Cameroonian plants against Gram-positive and Gram-negative multidrug resistant phenotypes. BMC Complement Altern Med. 16(1):388.
- Kuete V, Ngameni B, Wiench B, Krusche B, Horwedel C, Ngadjui BT, Efferth T. 2011. Cytotoxicity and mode of action of four naturally occuring flavonoids from the genus *Dorstenia*: gancaonin Q, 4-hydroxylonchocarpin, 6-prenylapigenin, and 6,8diprenyleriodictyol. *Planta Med.* 77(18):1984-1989.
- Tchinda CF, Voukeng KI, Beng VP, Kuete V. 2016. Antibacterial activities of the methanol extracts of *Albizia adianthifolia*, *Alchornea laxiflora*, *Laportea ovalifolia* and three other Cameroonian plants against multi-drug resistant Gram-negative bacteria *Saudi J Biol Sci*. 24:950-955.
- Mbaveng AT, Fotso GW, Ngnintedo D, Kuete V, Ngadjui BT, Keumedjio F, Andrae-Marobela K, Efferth T. 2018. Cytotoxicity of epunctanone and four other phytochemicals isolated from the medicinal plants *Garcinia epunctata* and *Ptycholobium contortum* towards multi-factorial drug resistant cancer cells. *Phytomedicine*. 48:112-119.
- Kuete V, Dzotam JK, Voukeng IK, Fankam AG, Efferth T. 2016. Cytotoxicity of methanol extracts of Annona muricata, Passiflora edulis and nine other Cameroonian medicinal plants towards multi-factorial drug-resistant cancer cell lines. Springerplus. 5(1):1666.
- Kuete V. 2013. Medicinal Plant Research in Africa: Pharmacology and Chemistry. Edited by Kuete V, 1 edn. Oxford: Elsevier. https://doi.org/10.1016/C2012-0-03354-6.
- Guefack MGF, Damen F, Mbaveng AT, Tankeo SB, Bitchagno GTM, Çelik İ, Simo Mpetga JD, Kuete V. 2020. Cytotoxic constituents of the bark of *Hypericum* roeperianum towards multidrug-resistant cancer cells. *Evidence-Based Complement Altern Med.* 2020:4314807.
- Kuete V, Mbaveng AT, Sandjo LP, Zeino M, Efferth T. 2017. Cytotoxicity and mode of action of a naturally occurring naphthoquinone, 2-acetyl-7-methoxynaphtho[2,3b]furan-4,9-quinone towards multi-factorial drug-resistant cancer cells. *Phytomedicine*. 33:62-68.
- Omosa LK, Midiwo JO, Mbaveng AT, Tankeo SB, Seukep JA, Voukeng IK, Dzotam JK, Isemeki J, Derese S, Omolle RA, Efferth T, Kuete V. 2016. Antibacterial activity and structure-activity relationships of a panel of 48 compounds from kenyan plants against multidrug resistant phenotypes. *SpringerPlus*. 5:901.
- Kuete V, Sandjo LP, Kwamou GM, Wiench B, Nkengfack AE, Efferth T. 2014. Activity of three cytotoxic isoflavonoids from Erythrina excelsa and Erythrina senegalensis (neobavaisoflavone, sigmoidin H and isoneorautenol) toward multifactorial drug resistant cancer cells. *Phytomedicine*. 21(5):682-688.
- Kuete V, Sandjo LP, Mbaveng AT, Seukep JA, Ngadjui BT, Efferth T. 2015. Cytotoxicity of selected Cameroonian medicinal plants and *Nauclea pobeguinii* towards multi-factorial drug-resistant cancer cells. *BMC Complement Altern Med.* 15:309.
- Mbaveng AT, Bitchagno GTM, Kuete V, Tane P, Efferth T. 2019. Cytotoxicity of ungeremine towards multi-factorial drug resistant cancer cells and induction of apoptosis, ferroptosis, necroptosis and autophagy. *Phytomedicine*. 60:152832.
- Kuete V, Tangmouo JG, Penlap Beng V, Ngounou FN, Lontsi D. 2006. Antimicrobial activity of the methanolic extract from the stem bark of Tridesmostemon omphalocarpoides (Sapotaceae). J Ethnopharmacol. 104(1-2):5-11.

- Kuete V, Efferth T. 2010. Cameroonian medicinal plants: pharmacology and derived natural products. Front Pharmacol. 1:123.
- Mbaveng AT, Kuete V, Efferth T. 2017. Potential of Central, Eastern and Western Africa medicinal plants for cancer therapy: spotlight on resistant cells and molecular targets. *Front Pharmacol.* 8:343.
- Kuete V, Efferth T. 2015. African flora has the potential to fight multidrug resistance of cancer. *BioMed Res Int.* 2015:914813.
- Kuete V. 2023. Chapter Twelve Ethnopharmacology, phytochemistry and pharmacology of potent antibacterial medicinal plants from Africa. Advances in Botanical Research, 107:353-660. https://doi.org/10.1016/bs.abr.2022.08.022.

30.

- http://www.llifle.com/Encyclopedia/SUCCULENTS/Family/Crassulaceae/290 88/Kalanchoe crenata: Kalanchoe crenata (Andrews) Haw. Accessed on January 22, 2023.
- 31. Sofowora A. 1993. Medical plants and traditional medicine in Africa. Ibadan: Spectrum books Ltd.
- Kuete V, Fokou FW, Karaosmanoğlu O, Beng VP, Sivas H. 2017. Cytotoxicity of the methanol extracts of *Elephantopus mollis*, *Kalanchoe crenata* and 4 other Cameroonian medicinal plants towards human carcinoma cells. *BMC Complement Altern Med*. 17(1):280.
- Nguelefack TB, Nana P, Atsamo AD, Dimo T, Watcho P, Dongmo AB, Tapondjou LA, Njamen D, Wansi SL, Kamanyi A. 2006. Analgesic and anticonvulsant effects of extracts from the leaves of *Kalanchoe crenata* (Andrews) Haworth (Crassulaceae). J *Ethnopharmacol.* 106(1):70-75.
- Kamgang R, Mboumi RY, Fondjo AF, Tagne MA, N'Dille G P, Yonkeu JN. 2008. Antihyperglycaemic potential of the water-ethanol extract of *Kalanchoe crenata* (Crassulaceae). J Nat Med. 62(1):34-40.
- Kamgang R, Foyet AF, Essame JL, Ngogang JY. 2012. Effect of methanolic fraction of *Kalanchoe crenata* on metabolic parameters in adriamycin-induced renal impairment in rats. *Indian J Pharmacol.* 44(5):566-570.
- Akinsulire OR, Aibinu IE, Adenipekun T, Adelowotan T, Odugberni T. 2007. In vitro antimicrobial activity of crude extracts from plants Bryophyllum pinnatum and Kalanchoe crenata. Afr J Trad Complement Altern Med. 4(3):338-344.
- Mapie Tiwa S, Matieta VY, Ngakam R, Kengne Fonkou G, Megaptche JF, Nayim P, Mbaveng AT, Kuete V. 2024. Antibacterial potential and modes of action of methanol extracts of *Elephantopus mollis* Kunth (Asteraceae) against multidrugresistant Gram-negative bacteria overexpressing efflux pumps. *Invest Med Chem Pharmacol.* 7(1):86.
- Ngakam R, Matieta VY, Kengne Fonkou G, Mapie Tiwa S, Megaptche JF, Nayim P, Mbaveng AT, Kuete V. 2024. Antibacterial potential and modes of action of methanol extracts of flowers and leaves of *Vernonia glabra* (Steetz) Vatke (Asteraceae) against multidrug-resistant Gram-negative bacteria overexpressing efflux pumps. *Invest Med Chem Pharmacol.* 7(1):87.
- Kengne Fonkou G, Matieta VY, Mapie Tiwa S, Ngakam R, Megaptche JF, Nayim P, Kuete V, Mbaveng AT. 2024. Botanicals from *Aframomum letestuanum* Gagnep. (Zingiberaceae) can overcome the multidrug resistance of Klebsiella species overexpressing AcrAB-ToIC efflux pumps. *Invest Med Chem Pharmacol.* 7(1):88.
- Kuete V, Alibert-Franco S, Eyong KO, Ngameni B, Folefoc GN, Nguemeving JR, Tangmouo JG, Fotso GW, Komguem J, Ouahouo BMW *et al.* 2011. Antibacterial activity of some natural products against bacteria expressing a multidrug-resistant phenotype. *Int J Antimicrob Agents*. 37(2):156-161.
- Kuete V, Ngameni B, Tangmouo JG, Bolla JM, Alibert-Franco S, Ngadjui BT, Pages JM. 2010. Efflux pumps are involved in the defense of Gram-negative bacteria against the natural products isobavachalcone and diospyrone. *Antimicrob Agents Chemother*. 54(5):1749-1752.
- Chollet R, Chevalier J, Bryskier A, Pagès J-M. 2004. The AcrAB-ToIC pump is involved in macrolide resistance but not in telithromycin efflux in Enterobacter aerogenes and Escherichia coli. ntimicrob Agents Chemother. 48(9):3621-3624.
- Dzotam JK, Touani FK, Kuete V. 2016. Antibacterial activities of the methanol extracts of *Canarium schweinfurthii* and four other Cameroonian dietary plants against multi-drug resistant Gram-negative bacteria. *Saudi J Biol Sci.* 23:565-570.
- Lorenzi V, Muselli A, Bernardini AF, Berti L, Pages JM, Amaral L, Bolla JM. 2009. Geraniol restores antibiotic activities against multidrug-resistant isolates from Gramnegative species. *Antimicrob Agents Chemother*. 53(5):2209-2211.
- 45. Dzotam JK, Simo IK, Bitchagno G, Celik I, Sandjo LP, Tane P, Kuete V. 2018. In vitro antibacterial and antibiotic modifying activity of crude extract, fractions and 3',4',7-trihydroxyflavone from Myristica fragrans Houtt against MDR Gram-negative enteric bacteria. BMC Complement Altern Med. 18(1):15.
- Seukep JA, Sandjo LP, Ngadjui BT, Kuete V. 2016. Antibacterial and antibioticresistance modifying activity of the extracts and compounds from *Nauclea* pobeguinii against Gram-negative multi-drug resistant phenotypes. *BMC Complement Altern Med.* 16:193.
- Dzotam JK, Kuete V. 2017. Antibacterial and antibiotic-modifying activity of methanol extracts from six cameroonian food plants against multidrug-resistant enteric bacteria. *BioMed Res Int.* 2017:1583510.
- Ghisalberti D, Masi M, Pages JM, Chevalier J. 2005. Chloramphenicol and expression of multidrug efflux pump in *Enterobacter aerogenes*. *Biochem Biophys Res Commun*. 328(4):1113-1118.
- Mallea M, Chevalier J, Bornet C, Eyraud A, Davin-Regli A, Bollet C, Pages JM. 1998. Porin alteration and active efflux: two in vivo drug resistance strategies used by *Enterobacter aerogenes*. *Microbiology*. 144 (Pt 11):3003-3009.
- Seukep JA, Fankam AG, Djeussi DE, Voukeng IK, Tankeo SB, Noumdem JA, Kuete AH, Kuete V. 2013. Antibacterial activities of the methanol extracts of seven Cameroonian dietary plants against bacteria expressing MDR phenotypes. *Springerplus*. 2:363.

- Tran QT, Mahendran KR, Hajjar E, Ceccarelli M, Davin-Regli A, Winterhalter M, Weingart H, Pages JM. 2010. Implication of porins in beta-lactam resistance of Providencia stuartii. J Biol Chem. 285(42):32273-32281.
- Touani FK, Seukep AJ, Djeussi DE, Fankam AG, Noumedem JA, Kuete V. 2014. Antibiotic-potentiation activities of four Cameroonian dietary plants against multidrug-resistant Gram-negative bacteria expressing efflux pumps. BMC Complement Altern Med. 14:258.
- Voukeng IK, Kuete V, Dzoyem JP, Fankam AG, Noumedem JA, Kuiate JR, Pages JM. 2012. Antibacterial and antibiotic-potentiation activities of the methanol extract of some Cameroonian spices against Gram-negative multi-drug resistant phenotypes. *BMC Res Notes*. 5:299.
- Fankam AG, Kuiate JR, Kuete V. 2015. Antibacterial and antibiotic resistance modifying activity of the extracts from *allanblackia gabonensis, combretum molle* and *gladiolus quartinianus* against Gram-negative bacteria including multi-drug resistant phenotypes. *BMC Complement Altern Med.* 15:206.
- Kuete V, Betrandteponno R, Mbaveng AT, Tapondjou LA, Meyer JJ, Barboni L, Lall N. 2012. Antibacterial activities of the extracts, fractions and compounds from Dioscorea bulbifera. BMC Complement Altern Med. 12:228.
- Mbaveng AT, Sandjo LP, Tankeo SB, Ndifor AR, Pantaleon A, Nagdjui BT, Kuete V. 2015. Antibacterial activity of nineteen selected natural products against multi-drug resistant Gram-negative phenotypes. *Springerplus*. 4:823.
- Noumedem JA, Mihasan M, Kuiate JR, Stefan M, Cojocaru D, Dzoyem JP, Kuete V. 2013. *In vitro* antibacterial and antibiotic-potentiation activities of four edible plants against multidrug-resistant Gram-negative species. *BMC Complement Altern Med.* 13:190.
- Cox SD, Mann CM, Markham JL, Bell HC, Gustafson JE, Warmington JR, Wyllie SG. 2000. The mode of antimicrobial action of the essential oil of *Melaleuca alternifolia* (tea tree oil). *J Appl Microbiol.* 88(1):170-175.
- Guefack M-GF, Messina NDM, Mbaveng AT, Nayim P, Kuete JRN, Matieta VY, Chi GF, Ngadjui BT, Kuete V. 2022. Antibacterial and antibiotic-potentiation activities of the hydro-ethanolic extract and protoberberine alkaloids from the stem bark of *Enantia chlorantha* against multidrug-resistant bacteria expressing active efflux pumps. J Ethnopharmacol. 296:115518.
- Manekeng HT, Mbaveng AT, Nguenang GS, Seukep JA, Wamba BEN, Nayim P, Yinkfu NR, Fankam AG, Kuete V. 2018. Anti-staphylococcal and antibioticpotentiating activities of seven Cameroonian edible plants against resistant phenotypes. *Invest Med Chem Pharmacol.* 1:7.
- Kovač J, Gavarić N, Bucar F, Smole Možina S. 2014. Antimicrobial and resistance modulatory activity of Alpinia katsumadai seed phenolic extract, essential oil and post-distillation extract. *Food Technol Biotechnol.* 52(2):248-254.
- Kuete V. 2010. Potential of Cameroonian plants and derived products against microbial infections: a review. *Planta Med.* 76(14):1479-1491.
- Kuete V, 2023. Chapter Six Potential of African medicinal plants against Enterobacteria: Classification of plants antibacterial agents. Advances in Botanical Research. 106: 151-335. <u>https://doi.org/10.1016/bs.abr.2022.1008.1006</u>.
- Tankeo SB, Kuete V. 2023. Chapter Seven African plants acting on Pseudomonas aeruginosa: Cut-off points for the antipseudomonal agents from plants. Advances in Botanical Research. 106: 337-412. https://doi.org/10.1016/bs.abr.2022.08.007.
- Mims C, Playfair J, Roitt I, Wakelin D, Williams R. 1993. Antimicrobials and chemotherapy. In: Mims CA, et al Eds, Med Microbiol Rev. 35:1-34.
- Nayim P, Mbaveng AT, Wamba BEN, Fankam AG, Dzotam JK, Kuete V. 2018. Antibacterial and antibiotic-potentiating activities of thirteen Cameroonian edible plants against gram-negative resistant phenotypes. *ScientificWorldJournal*. 2018:4020294.
- Tchana ME, Fankam AG, Mbaveng AT, Nkwengoua ET, Seukep JA, Tchouani FK, Nyassé B, Kuete V. 2014. Activities of selected medicinal plants against multi-drug resistant Gram-negative bacteria in Cameroon. *Afr Health Sci.* 14(1):167-172.
- Mbaveng AT, Kuete V, Nguemeving JR, Beng VP, Nkengfack AE, Marion Meyer JJ, Lall N, Krohn K. 2008. Antimicrobial activity of the extracts and compounds from Vismia guineensis (Guttiferae). Asian Journal of Traditional Medicine. 3:211-223.
- Mbaveng AT, Kuete V, Mapunya BM, Beng VP, Nkengfack AE, Meyer JJ, Lall N. 2011. Evaluation of four Cameroonian medicinal plants for anticancer, antigonorrheal and antireverse transcriptase activities. *Environ Toxicol Pharmacol.* 32(2):162-167.
- Kuete V, Simo IK, Ngameni B, Bigoga JD, Watchueng J, Kapguep RN, Etoa FX, Tchaleu BN, Beng VP. 2007. Antimicrobial activity of the methanolic extract, fractions and four flavonoids from the twigs of *Dorstenia angusticomis* Engl. (Moraceae). *J Ethnopharmacol.* 112(2):271-277.
- Kuete V, Metuno R, Ngameni B, Tsafack AM, Ngandeu F, Fotso GW, Bezabih M, Etoa F-X, Ngadjui BT, Abegaz BM et al. 2007. Antimicrobial activity of the methanolic extracts and compounds from *Treculia obovoidea* (Moraceae). J Ethnopharmacol. 112(3):531-536.
- Nielsen TR, Kuete V, Jager AK, Meyer JJ, Lall N: Antimicrobial activity of selected South African medicinal plants. *BMC Complement Altern Med* 2012, 12:74.
- Kuete V, Ngameni B, Mbaveng AT, Ngadjui B, Meyer JJ, Lall N. 2010. Evaluation of flavonoids from *Dorstenia barteri* for their antimycobacterial, antigonorrheal and antireverse transcriptase activities. *Acta Trop.* 116(1):100-104.
- Nono EC, Mkounga P, Kuete V, Marat K, Hultin PG, Nkengfack AE. 2010. Pycnanthulignenes A-D, antimicrobial cyclolignene derivatives from the roots of *Pycnanthus angolensis. J Nat Prod.* 73(2):213-216.
- Komguem J, Meli AL, Manfouo RN, Lontsi D, Ngounou FN, Kuete V, Kamdem HW, Tane P, Ngadjui BT, Sondengam BL *et al.* 2005. Xanthones from *Garcinia smeathmannii* (Oliver) and their antimicrobial activity. *Phytochemistry*. 66(14):1713-1717.

- Kuete V, Tangmouo JG, Marion Meyer JJ, Lall N. 2009. Diospyrone, crassiflorone and plumbagin: three antimycobacterial and antigonorrhoeal naphthoquinones from two *Diospyros* spp. *Int J Antimicrob Agents*. 34(4):322-325.
- Tekwu EM, Askun T, Kuete V, Nkengfack AE, Nyasse B, Etoa FX, Beng VP. 2012. Antibacterial activity of selected Cameroonian dietary spices ethno-medically used against strains of *Mycobacterium tuberculosis. J Ethnopharmacol.* 142(2):374-382.
- Nguemeving JR, Azebaze AG, Kuete V, Eric Carly NN, Beng VP, Meyer M, Blond A, Bodo B, Nkengfack AE. 2006. Laurentixanthones A and B, antimicrobial xanthones from *Vismia laurentii*. *Phytochemistry*. 67(13):1341-1346.
- 79. Kuete V, Sandjo LP. 2012. Isobavachalcone: an overview. Chin J Integr Med. 18(7):543-547.
- Kuete V, Wansi JD, Mbaveng AT, Kana Sop MM, Tadjong AT, Beng VP, Etoa FX, Wandji J, Meyer JJM, Lall N. 2008. Antimicrobial activity of the methanolic extract and compounds from *Teclea afzelii* (Rutaceae). S Afr J Bot. 74(4):572-576.
- Ngameni B, Kuete V, Simo IK, Mbaveng AT, Awoussong PK, Patnam R, Roy R, Ngadjui BT. 2009. Antibacterial and antifungal activities of the crude extract and compounds from *Dorstenia turbinata* (Moraceae). S Afr J Bot. 75(2):256-261.

- Kuete V, Tangmouo JG, Penlap Beng V, Ngounou FN, Lontsi D. 2006. Antimicrobial activity of the methanolic extract from the stem bark of *Tridesmosternon* omphalocarpoides (Sapotaceae). J Ethnopharmacol. 104(1–2):5-11.
- Ngounou FN, Manfouo RN, Tapondjou LA, Lontsi D, Kuete V, Penlap V, Etoa FX, Dubois MAL, Sondengam BL. 2005. Antimicrobial diterpenoid alkaloids from *Erythrophleum suaveolens* (guill. & perr.) brenan. *Bull Chem Soc Ethiop*. 19(2):221-226.
- Jepkoech C, Omosa LK, Nchiozem-Ngnitedem VA, Kenanda EO, Guefack MF, Mbaveng AT, Kuete V, Heydenreich M. 2021. Antibacterial secondary metabolites from Vernonia auriculifera Hiern (Asteraceae) against MDR phenotypes. *Nat Prod Res.* 36(12):3203-3206.
- Braga LC, Leite AA, Xavier KG, Takahashi JA, Bemquerer MP, Chartone-Souza E, Nascimento AM. 2005. Synergic interaction between pomegranate extract and antibiotics against Staphylococcus aureus. Can J Microbiol. 51(7):541-547.
- Fankam AG, Kuiate JR, Kuete V. 2017. Antibacterial and antibiotic resistance modulatory activities of leaves and bark extracts of *Recinodindron heudelotii* (Euphorbiaceae) against multidrug-resistant Gram-negative bacteria. *BMC Complement Altern Med.* 17(1):168.